

Seasonal Variation of Urushiol Content in Poison Oak Leaves

Barbara L. Gartner, Christian Wasser, Eloy Rodriguez, and William L. Epstein

Urushiol is the causative agent in poison oak and poison ivy dermatitis. Urushiol concentration in poison oak (*Toxicodendron diversilobum*) leaves was quantified weekly between mid-August and late November for several individual genotypes of plants that had been grown in either sun, moderate shade, or deep shade. These individual plants were grown in the same garden to reduce variation caused by environmental factors. Plants cloned from a coastal genotype had higher levels of urushiol than those from an inland site, suggesting that people who live in coastal areas

may be at a greater risk for contracting poison oak dermatitis than people who live farther inland. The concentration of urushiol remained relatively constant in the inland clones until early November, at which point it increased by three to six times. This increase in urushiol may be responsible for the greater number of cases that some dermatologists have noticed in the autumn. The level of urushiol in leaf litter was very low, and so it can be considered nonallergenic.

Copyright © 1993 by W.B. Saunders Company

PLANTS of the genus *Toxicodendron* (formerly known as *Rhus* Anacardiaceae) are notorious as causes of contact dermatitis.^{1,2} The dermatitis produced by these plants is caused by the urushiol fraction, which is primarily composed of 3-pentadecylcatechol and three heptadecylcatechol analogs in poison oak.^{3,4} Several methods have been developed to lessen the severity of the rashes produced by urushiol⁵ but the best precaution is to avoid contact with the plant and the chemicals in it.

There are four species in this genus in North America, but *Toxicodendron diversilobum* and *T. radicans* are the most common, and thus cause the most cases of dermatitis. The only unambiguous names for members of this genus are the scientific names because "poison oak" and "poison ivy" are used to indicate the same species in some regions and different species between regions. *T. radicans* (often called poison ivy) is the predominant species in the eastern half of the United States, and it is divided into a number of subspecies⁶; *T. rydbergii* has a wide range in the middle of the United States, and *T. toxicarium* is found in the Southeast (Fig 1).

The subject of this paper, *T. diversilobum* (most often referred to as poison oak), is the only species in this genus to live west of the Sierra Nevada (Fig 1). It will be referred to hereafter as poison oak. Poison oak can assume a continuum of growth forms from a short shrub to a 30-meter-tall vine. It can be found in full sun or deep shade, in creek beds, on dry hillsides, on a variety of soils, on windswept coastal bluffs, and in a variety of other habitats: coniferous forest, chaparral, and meadows.⁷ The differences between the vine and shrub growth forms have recently been shown to be

caused by environmental factors (the presence or absence of physical support) and not heredity.⁸

Many studies have been performed on the variability of the chemical constituents of plants. A cursory examination of recent literature indicates that chemical constituents such as monoterpenes and flavonoids, vary markedly in pine trees from different geographic regions.⁹⁻¹¹ Different plants also have been studied for seasonal and environmental variability in the content of monoterpenes,¹² alkaloids,¹³ and furanocoumarins.¹⁴ Studies have established that chemical constituents such as furanocoumarins, can increase after the stress of fungal infections or cold night temperatures.¹⁵

The comparative urushiol composition and content in different species of *Toxicodendron* have been extensively studied, for example, references 16 and 17, and Baer et al¹⁸ established that the urushiol content of leaves can vary with the position of the leaf on the stem. However, no studies have been reported on the differences in concentration and composition of urushiol of plants with different growth habits. Moreover, no data are available on the seasonal variation in urushiol content of poison oak.

From the Department of Biological Sciences, Stanford University, Stanford, CA; the Department of Forest Products, Forest Research Lab 105, Oregon State University, Corvallis, OR; Phytochemistry and Toxicology Laboratory, Department of Developmental and Cell Biology, University of California, Irvine, CA; and School of Medicine, Department of Dermatology, University of California, San Francisco, CA.

Address reprint requests to Barbara L. Gartner, MD, Department of Forest Products, Forest Research Lab 105, Oregon State University, Corvallis, OR 97331.

Copyright © 1993 by W.B. Saunders Company
1046-199X/93/0401-0006\$03.00/0

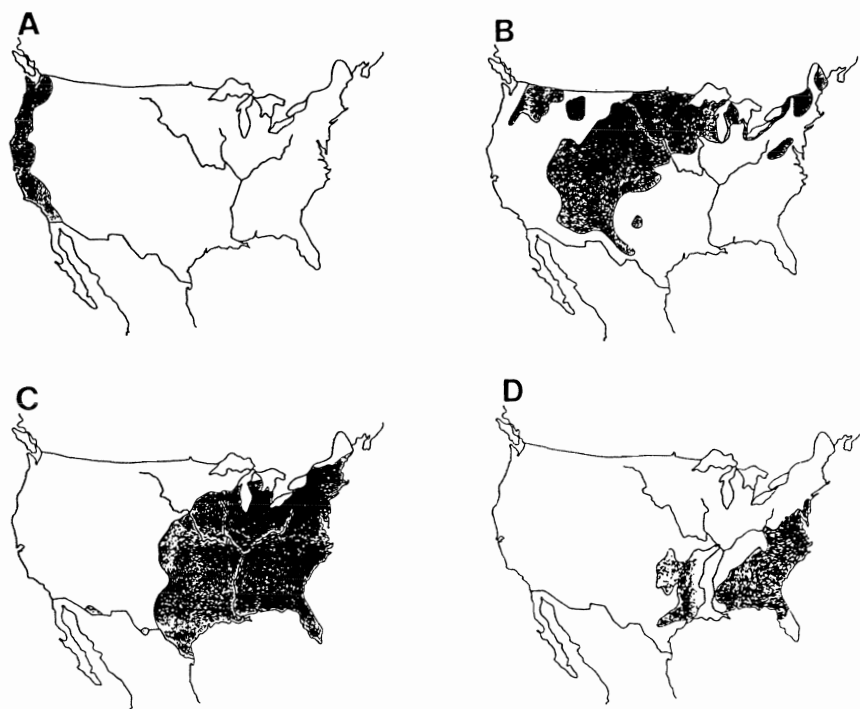


Fig 1. Geographic ranges of the four species of *Toxicodendron* in the United States: (A) *T. diversilobum* (Western poison oak); (B) *T. rydbergii* (Rydberg's poison ivy); (C) *T. radicans* (poison ivy); and (D) *T. toxicarium* (Eastern poison oak). Data from Gillis.⁶

MATERIALS AND METHODS

Collection of Samples

Poison oak plants were grown from cuttings taken from three different source plants, a vine (plant A) and a shrub (plant B) from Jasper Ridge Biological Preserve, Woodside, CA, in the coastal range near Stanford University, and a shrub (plant C) from a bluff overlooking Pomponio Beach, CA, 20 km southwest of Jasper Ridge. Three replicate cuttings ("clones") from each source plant were planted into each of three light environments: full sun, 45% sun, and 15% sun.¹⁹ Clones were sampled for this study 1½ years later, to assure that all buds and leaves had developed in the light environment in which the clone, not the source plant, had grown.

On May 14, 1989, samples were collected from all plants to learn the variance among source plants and light treatments for total urushiol and the proportion of the different catechols in the urushiol. Each sample consisted of nine leaves (three from each of the three clones of a given plant). Samples were stored in plastic bags and analyzed within 4 days of collection.

From mid-August 1989 until late November when the majority of leaves had dropped, clones A and B in 15% sun were

sampled weekly to determine the seasonal change in total urushiol content of the leaves. Three weeks later, their abscised (dead or fallen) leaves were sampled. We refer to these as "leaf litter."

Analysis of Samples

Poison oak leaves (2.0 g, fresh weight) were extracted with 30 mL of methanol. The extracts were then centrifuged for 5 minutes at 3,000 rpm. Five milliliters of the supernatant was evaporated to dryness, redissolved in 5 mL of dichloromethane, filtered, and evaporated. The extract was then redissolved in 5 mL of high-pressure liquid chromatography (HPLC)-grade methanol. Each sample was then filtered through a Miniclean Cartridge (C18, Alltech, Deerfield, IL) before injection on the HPLC column. HPLC analysis was performed using a C18 reversed phase cartridge (Alltech, Econosphere C18, 250 × 4.6 mm, 5 mm particle size) and methanol-water (95:5) mixture at 1.2 mL/min with UV detection at 275 nm.

Four different catechol derivatives (Fig 2) were detected, but heptadecylcatechol was present in extremely low concentrations and could not be analyzed accurately. Heptadecylcate-

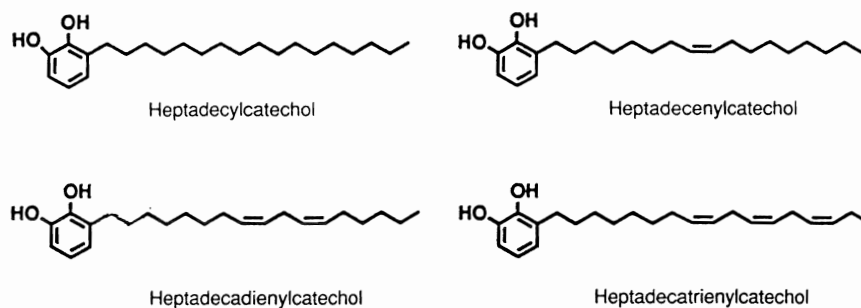


Fig 2. Catechol derivatives of poison oak: heptadecylcatechol, heptadecenylcatechol (HDEC), heptadecadienylcatechol (HDDC), and heptadecatrienylcatechol (HDTTC).

chol (HDEC), heptadecadienylcatechol (HDDC), and heptadecatrienylcatechol (HDTC) were present in larger concentrations and were analyzed throughout the experiment. The concentration of catechols in each sample was determined by comparison with a set of external standards of purified urushiol (0.1, 0.2, 0.3, 0.4, and 0.5 mg/mL). The detector response was linear over the range of these standards. Urushiol used for standards was purified according to a procedure described elsewhere.²⁰

RESULTS AND DISCUSSION

On May 14, 1989, after leaves had fully expanded but before they had begun to senesce, the foliar concentration of urushiol was greater in clones of plants from the coastal area (plant C) than in the clones from Jasper Ridge (plants A and B, Fig 3; significant at $P < .01$, one-factor analysis of variance [ANOVA]). The individual plants that grew in full sun had slightly more urushiol than the individual plants that grew in shaded areas for all clones. A lower proportion of the urushiol was composed of HDTC in individual plants grown in the full sun rather than in individual plants grown in the shade.

The total foliar urushiol content was also analyzed 17 times between late spring and late fall in the clones from plants A and B (Fig 4). The amount of urushiol remained relatively constant until early November, at which point it increased from the baseline level (about 0.02% to 0.4% by fresh weight) to 0.9% to 2.6% of fresh weight. High content was maintained for 2 weeks, after which the leaves showed low levels of urushiol again. Two

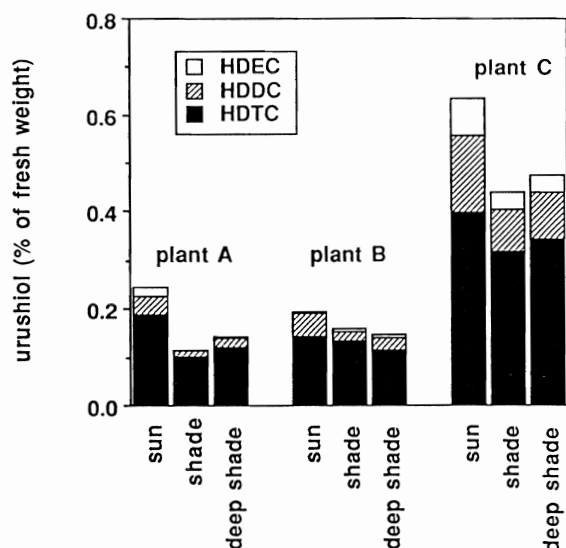


Fig 3. Foliar urushiol content and composition in three clones (from an inland vine [A], and inland shrub [B], and a coastal shrub [C]), in three different light environments.

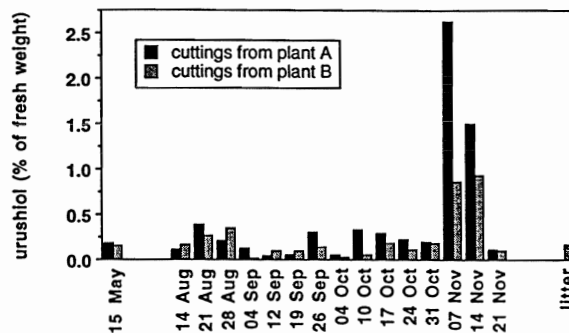


Fig 4. Seasonal variation in foliar urushiol content in clones of two plants growing in deep shade.

weeks after the increase in urushiol, the leaves abscised (dropped). The leaf litter contained a very small quantity of urushiol. These data were collected on a fresh weight basis, and there are many more dead than live leaves per gram; therefore, the low level of urushiol in dead leaves underemphasizes the actual decrease in urushiol concentration that occurs between live and dead leaves. Thus, if urushiol content were expressed per leaf surface area rather than per fresh weight, the contrast between urushiol content of litter and attached leaves would be greater.

These results led us to conclude that urushiol concentration and composition are affected by the light environment of the plant. The greater concentration of urushiols in the individual plants cloned from plant C (the coastal shrub) suggested that a plant's "genotypic origin" or "race" can affect urushiol content. The increase in catechol concentration observed during November occurred as the leaves senesced, shortly before they fell off the plant. This could be a byproduct of the stresses of cold nights, but more likely, the increase is caused by an apparent enrichment in urushiols as other components (carbon, nutrients, and water) are exported from the leaf. Up to 30% of a leaf's carbon and 70% of its nitrogen and phosphorus can be exported from a leaf before senescence,²¹ and the leaf can lose water as well; therefore, its fresh weight per unit leaf area would be expected to be much lower in litter than living tissue.

CONCLUSIONS

This study suggests that the environment and the plant's genetic composition (genotype) influence the quantity and composition of urushiols produced in *T. diversilobum*. Urushiol is a threat to the health of more than 50% of the North Ameri-

can population,²² so it is essential to identify when and where the risk of contacting these allergens is highest. As long as the leaves are attached to the plant, their urushiol content is sufficient to cause an allergic reaction in sensitized individuals. However, the leaf litter can be considered benign. Our study suggests that the chances of contracting poison oak dermatitis are higher in coastal areas and in late fall as the leaves begin to senesce. Many dermatolo-

gists have noted an increase in poison oak cases in the fall, but its basis had not been investigated previously.

ACKNOWLEDGMENT

We thank H.A. Mooney for discussion, J. West for critically reviewing the manuscript, and the Department of Plant Biology of the Carnegie Institution of Washington at Stanford University for permitting us to grow poison oak on their grounds.

REFERENCES

1. Horsfield T: An experimental dissertation on the *Rhus vernix*, *Rhus radicans*, and *Rhus glabrum*: Commonly known in Pennsylvania by the names of poison-ash, poison-vine, and common sumac. Doctoral thesis, University of Pennsylvania, Philadelphia, PA, 1798
2. Epstein WL, Baer H, Dawson CR, et al: Poison oak hypersensitization: Evaluation of purified urushiol. *Arch Dermatol* 109:356-360, 1976
3. Dawson CR: The chemistry of poison ivy. *Trans NY Acad Sci* 18:427-443, 1956
4. Corbett MD, Billets S: Characterization of poison oak urushiol. *J Pharm Sci* 64:1715-1718, 1975
5. Epstein WL: Topical prevention of poison ivy/oak dermatitis. *Arch Dermatol* 125:449-501, 1982
6. Gillis WT: The systematics and ecology of poison-ivy and the poison-oaks (*Toxicodendron*, Anacardiaceae). *Rhodora* 73:72-160, 161-237, 370-443, 465-540, 1971
7. Jepson WL: *Rhus diversiloba* T. & G., in A Flora of California, vol 2. San Francisco, CA, California School Book Depository, 1936, pp 445-446
8. Gartner BL: Is the climbing habit of poison oak ecotypic? *Funct Ecol* 5:696-704, 1991
9. Zavarin E, Rafii Z, Cool LG, et al: Geographic monoterpene variability of *Pinus albicaulis*. *Biochem Syst Ecol* 19:147-156, 1991
10. Lauranson J, Lebreton P: Flavonoid variability within and between natural populations of *Pinus uncinata*. *Biochem Syst Ecol* 19:659-664, 1991
11. Zavarin E, Snajberk K, Cool L: Monoterpene variability of *Pinus monticola* wood. *Biochem Syst Ecol* 18:117-124, 1990
12. Mihaliak CH, Couvet D, Lincoln DE: Genetic and environmental contributions to variation in leaf mono- and sesquiterpenes of *Heterotheca subaxillaris*. *Biochem Syst Ecol* 7:529-533, 1989
13. Bernáth J, Dános B, Veres T, et al: Variation in alkaloid production in poppy ecotypes: Responses to different environments. *Biochem Syst Ecol* 16:171-178, 1988
14. Zobel AM, Brown SA: Seasonal changes of furanocoumarin concentrations in leaves of *Heracleum lanatum*. *J Chem Ecol* 16:1623-1634, 1990
15. Beier RC, Oertli EH: Psoralen and other linear furocoumarins as phytoalexins in celery. *Phytochemistry* 22:2595-2597, 1983
16. Billets S, Craig JC, Corbett MD, et al: Analysis of urushiol content of poison ivy and poison oak. *Phytochemistry* 15:533-535, 1976
17. Gross M, Baer H, Fales HM: Urushiols of poisonous Anacardiaceae. *Phytochemistry* 14:2263-2266, 1975
18. Baer H, Hooton M, Fales H, et al: Catecholic and other constituents of the leaves of *Toxicodendron radicans* and variation of urushiol concentrations within one plant. *Phytochemistry* 19:799-802, 1980
19. Gartner BL: Structural stability and architecture of vines vs. shrubs of poison oak, *Toxicodendron diversilobum*. *Ecology* 72:2005-2015, 1991
20. Dupuis G: Studies on poison ivy. In vitro lymphocyte transformation by urushiol-protein conjugates. *Br J Dermatol* 101:617-624, 1979
21. Bloom AJ, Chapin FS III, Mooney HA: Resource limitation in plants—An economic analogy. *Annu Rev Ecol Syst* 16:363-392, 1985
22. Epstein WL: The poison oak picker of Pennypack Park: The continuing saga of poison ivy. *J Invest Dermatol* 88:7s-11s, 1987 (suppl)

can population,²² so it is essential to identify when and where the risk of contacting these allergens is highest. As long as the leaves are attached to the plant, their urushiol content is sufficient to cause an allergic reaction in sensitized individuals. However, the leaf litter can be considered benign. Our study suggests that the chances of contracting poison oak dermatitis are higher in coastal areas and in late fall as the leaves begin to senesce. Many dermatolo-

gists have noted an increase in poison oak cases in the fall, but its basis had not been investigated previously.

ACKNOWLEDGMENT

We thank H.A. Mooney for discussion, J. West for critically reviewing the manuscript, and the Department of Plant Biology of the Carnegie Institution of Washington at Stanford University for permitting us to grow poison oak on their grounds.

REFERENCES

1. Horsfield T: An experimental dissertation on the *Rhus* *venix*, *Rhus radicans*, and *Rhus glabrum*: Commonly known in Pennsylvania by the names of poison-ash, poison-vine, and common sumac. Doctoral thesis, University of Pennsylvania, Philadelphia, PA, 1798
2. Epstein WL, Baer H, Dawson CR, et al: Poison oak hypersensitization: Evaluation of purified urushiol. *Arch Dermatol* 109:356-360, 1976
3. Dawson CR: The chemistry of poison ivy. *Trans NY Acad Sci* 18:427-443, 1956
4. Corbett MD, Billets S: Characterization of poison oak urushiol. *J Pharm Sci* 64:1715-1718, 1975
5. Epstein WL: Topical prevention of poison ivy/oak dermatitis. *Arch Dermatol* 125:449-501, 1982
6. Gillis WT: The systematics and ecology of poison-ivy and the poison-oaks (*Toxicodendron*, Anacardiaceae). *Rhodora* 73:72-160, 161-237, 370-443, 465-540, 1971
7. Jepson WL: *Rhus diversiloba* T. & G., in A Flora of California, vol 2. San Francisco, CA, California School Book Depository, 1936, pp 445-446
8. Gartner BL: Is the climbing habit of poison oak ecotypic? *Funct Ecol* 5:696-704, 1991
9. Zavarin E, Rafii Z, Cool LG, et al: Geographic monoterpene variability of *Pinus albicaulis*. *Biochem Syst Ecol* 19:147-156, 1991
10. Lauranson J, Lebreton P: Flavonoid variability within and between natural populations of *Pinus uncinata*. *Biochem Syst Ecol* 19:659-664, 1991
11. Zavarin E, Snajberk K, Cool L: Monoterpene variability of *Pinus monticola* wood. *Biochem Syst Ecol* 18:117-124, 1990
12. Mihaliak CH, Couvet D, Lincoln DE: Genetic and environmental contributions to variation in leaf mono- and sesquiterpenes of *Heterotheca subaxillaris*. *Biochem Syst Ecol* 7:529-533, 1989
13. Bernáth J, Dános B, Veres T, et al: Variation in alkaloid production in poppy ecotypes: Responses to different environments. *Biochem Syst Ecol* 16:171-178, 1988
14. Zobel AM, Brown SA: Seasonal changes of furanocoumarin concentrations in leaves of *Heracleum lanatum*. *J Chem Ecol* 16:1623-1634, 1990
15. Beier RC, Oertli EH: Psoralen and other linear furocoumarins as phytoalexins in celery. *Phytochemistry* 22:2595-2597, 1983
16. Billets S, Craig JC, Corbett MD, et al: Analysis of urushiol content of poison ivy and poison oak. *Phytochemistry* 15:533-535, 1976
17. Gross M, Baer H, Fales HM: Urushiol of poisonous Anacardiaceae. *Phytochemistry* 14:2263-2266, 1975
18. Baer H, Hooton M, Fales H, et al: Catecholic and other constituents of the leaves of *Toxicodendron radicans* and variation of urushiol concentrations within one plant. *Phytochemistry* 19:799-802, 1980
19. Gartner BL: Structural stability and architecture of vines vs. shrubs of poison oak, *Toxicodendron diversilobum*. *Ecology* 72:2005-2015, 1991
20. Dupuis G: Studies on poison ivy. In vitro lymphocyte transformation by urushiol-protein conjugates. *Br J Dermatol* 101:617-624, 1979
21. Bloom AJ, Chapin FS III, Mooney HA: Resource limitation in plants—An economic analogy. *Annu Rev Ecol Syst* 16:363-392, 1985
22. Epstein WL: The poison oak picker of Pennypack Park: The continuing saga of poison ivy. *J Invest Dermatol* 88:7s-11s, 1987 (suppl)